

Accepted Manuscript

Egg Yolk IgY Antibodies: a Therapeutic Intervention Against Group A Rotavirus in Calves

C. Vega, M. Bok, L. Saif, F. Fernandez, V. Parreño

PII: S0034-5288(15)30053-9
DOI: doi: [10.1016/j.rvsc.2015.09.005](https://doi.org/10.1016/j.rvsc.2015.09.005)
Reference: YRVSC 2953

To appear in:

Received date: 5 May 2015
Revised date: 1 September 2015
Accepted date: 4 September 2015

Please cite this article as: Vega, C., Bok, M., Saif, L., Fernandez, F., Parreño, V., Egg Yolk IgY Antibodies: a Therapeutic Intervention Against Group A Rotavirus in Calves, (2015), doi: [10.1016/j.rvsc.2015.09.005](https://doi.org/10.1016/j.rvsc.2015.09.005)

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22

**Egg Yolk IgY Antibodies:
a Therapeutic Intervention Against Group A Rotavirus in Calves**

C. Vega¹, M. Bok¹, L. Saif², F. Fernandez¹ and V. Parreño¹.

¹ Instituto de Virología, CICV y A - INTA, Argentina. ² Food Animal Health Research Program (FAHRP). The Ohio Agricultural Research and Development Center, The Ohio State University, USA.

*Corresponding author: Dr. Viviana Parreño, Instituto de Virología, CICV y A - INTA, Castelar. Phone/FAX: 54114481-3006. E-mail: parreno.viviana@inta.gob.ar - Postal address: CC 25 (1712) Castelar – Bs. As., Argentina.

1. ABSTRACT

2

3 Bovine group A rotavirus (RVA) is considered the major cause of diarrhea in intensively
4 reared neonatal calves. Chicken egg yolk antibodies (IgY) are efficient in protecting
5 neonatal calves from RVA diarrhea; however, it is unclear the value of this intervention
6 in calves once diarrhea has appeared. The aim of the present study was to evaluate the
7 application of RVA-specific IgY as a passive treatment in those cases. The experimental
8 groups were: **G1**= RVA-specific IgY treatment; **G2**= no Ab treatment; and **G3**=
9 colostrum deprived + no Ab treatment. IgY treatment significantly reduced virus
10 shedding, diarrhea duration and severity compared to G2 and G3 calves. However, it
11 caused a partial suppression of systemic Ab responses to RVA that could be associated
12 with less severe diarrhea. The oral treatment with IgY for 7 days was associated with
13 significantly higher antibody secreting cell responses in the calves compared with the
14 other groups of animals.

15

16 **Keywords:** bovine, rotavirus, passive immunity, chicken antibodies, IgY.

17

2. INTRODUCTION

2
3 Numerous epidemiologic studies have shown that group A rotaviruses (RVA) are the
4 most frequent cause of severe gastroenteritis in young children and animals (Estes and
5 Kapikian, 2007; Dhama et al., 2009; Martella et al., 2010). In 1969, electron microscopy
6 was used to detect virus associated with neonatal calf diarrhea (Mebus et al., 1969).
7 Multiple etiological agents are responsible for neonatal diarrhea in calves including RVA,
8 Coronavirus, *Escherichia coli*, *Salmonella* species, *Clostridium perfringens*,
9 *Cryptosporidium* and *Coccidia* (Bellinzoni et al., 1990; Holland, 1990; Bendali et al.,
10 1999a; Svennerholm and Steele, 2004). Among these agents, bovine RVA is
11 considered the major cause of enteritis and diarrhea in artificially reared calves, causing
12 important economic losses related to death, treatment costs and reduction in weight
13 gain of affected animals (Malik et al., 1995; Martella et al., 2010). Diarrheal disease
14 severity is enhanced by several factors including herd management, environment and
15 host nutritional and immunological status (Saif and Smith, 1985; Bendali et al., 1999b).
16 The disease usually occurs in 2- to 8-week-old calves and the severity decreases as the
17 age increases. The incubation period is very short (12-24 hours) and the diarrhea is self-
18 limiting within 5 to 7 days, unless secondary bacterial infections occur (Svennerholm
19 and Steele, 2004; Dhama et al., 2009). Bovine RVA strains are genetically and
20 antigenically heterogeneous. Their genetic classification is based on the genes coding
21 the two outer capsid proteins, VP7 and VP4, determining G and P types, respectively.
22 The single most prevalent genotype combination among bovine RVA strains has been
23 G6P[5], although spatiotemporal and herd farm type (cattle or dairy) differences in RVA
24 strain distribution are observed worldwide (Garaicoechea et al., 2006; Badaracco et

1 al., 2012; Papp et al., 2013).

2 Bovine RVA is ubiquitous and infects the enterocytes of the tip of the villi within the
3 small intestine of neonatal calves. By multifactorial mechanisms, the infection disrupts
4 the efficiency of the absorptive enterocytes and potentially activates the enteric nervous
5 system, inducing a malabsorptive and secretory diarrhea (Bridger, 1994; Malik et al.,
6 1995; Lundgren and Svensson, 2001; Dhama et al., 2009; Martella et al., 2010). The
7 virus replicates quickly, generating high yields of virions and completing the
8 transmission cycle before the host immune mechanisms can intervene (Estes and
9 Kapikian, 2007). The affected calves manifest severe diarrhea, dehydration,
10 hyperthermia, anorexia and lethargy (Bellinzoni et al., 1990; Holland, 1990; Bendali et
11 al., 1999b; Dhama et al., 2009).

12 Good management and hygienic practices can help to reduce the severity of RVA
13 diarrhea in farm animals, while antibiotics can be used to control secondary bacterial
14 infections and fluid and electrolyte therapy can help to restore the fluid reserves,
15 minimizing calf mortality (Holland, 1990; Svennerholm and Steele, 2004). Being a
16 gastrointestinal pathogen, the local gut-associated lymphoid tissue (GALT) responses
17 are critical for virus clearance and prevention of RVA reinfection. The RVA-specific
18 passive maternal antibodies (Ab) acquired by colostrum intake (systemically absorbed
19 and re-secreted to the gut) as well as Ab present in maternal milk (acting locally) are key
20 factors in protecting the neonates (Stott and Fella, 1983; Besser et al., 1988; Saif and
21 Fernandez, 1996; Barrandeguy et al., 1998; Fernandez et al., 1998; Hodgins et al.,
22 1999; Kohara and Tsunemitsu, 2000; Parreño et al., 2004). Dams are immunized with
23 RVA-inactivated vaccines in the last third of pregnancy, between 60 to 30 days prior to
24 parturition to enhance maternal Ab levels in serum to promote a higher concentration of

1 IgG1 immunoglobulins in colostrum (Holland, 1990; Saif and Fernandez, 1996;
2 Barrandeguy et al., 1998; Svennerholm and Steele, 2004). Although Ab present in
3 colostrum enter the circulation, the protection against this enteric disease is primarily
4 dependent on the presence of RVA-specific Ab in the calves' intestinal lumen, indicating
5 that protection could be achieved by the presence of passive Ab in the gut lumen
6 combined with a rapid active antibody secreting cell (ASC) response in the intestinal
7 lamina propria (Saif and Fernandez, 1996; Fernandez et al., 1998). Regarding passive
8 immune protection, calves receiving pooled colostrum from vaccinated dams prior to gut
9 closure (within 4 hours after birth) were protected against severe diarrhea (Fernandez et
10 al., 1998; Parreño et al., 2004). However, RVA-specific Ab from colostrum decline after
11 calving and seem to be insufficient against high challenge doses of virus (Parreño et al.,
12 2004). On the other hand, supplementation of the milk diet with immune colostrum
13 significantly reduced diarrhea and delayed viral shedding onset (Kohara and
14 Tsunemitsu, 2000; Parreño et al., 2010). However, the production of immune milks
15 based on colostrum supplementation was not an industrially scalable alternative. This
16 highlights the need of another source of large amounts of Ab for immunotherapeutic
17 purposes in a cost-efficient way.

18 Recently, chickens have attracted considerable attention as an alternative source of
19 antibodies, as IgY is deposited in the egg yolk in large quantities, making chickens an
20 ideal source for specific polyclonal antibodies (Touchette et al., 2003; Kovacs-Nolan and
21 Mine, 2004). There are several reports of chicken egg yolk Ab (IgY) being highly
22 effective in protecting neonatal calves from bovine RVA diarrhea (Ikemori et al., 1992,
23 1997; Kuroki et al., 1994, 1997). Our group has described that prophylactic provision of
24 hyperimmune crude fluid egg yolk to newborn calves during the first 14 consecutive

1 days of life significantly reduced the severity of diarrhea after oral inoculation with
2 virulent bovine RVA. The treatment also prevented RVA diarrhea and virus shedding
3 after a second virus challenge (seven days after the end of the passive Ab treatment).
4 We observed that egg yolk treatment promoted a modulatory effect on ASC responses
5 in the gut, inducing high numbers of bovine RVA-specific IgA ASCs (Vega et al., 2011).
6 This 14-day-long prophylactic treatment could be advantageous in dairy herds where
7 bovine RVA is a major epidemiological concern. Still, other oral IgY-based treatments
8 can be useful in dairy farms where RVA-associated diarrhea is a concern but less
9 severe, decreasing the therapy's costs. Little is known about the usefulness of IgY Ab
10 as a treatment to bovine RVA once virus infection has already induced diarrhea under
11 controlled environmental conditions. The aim of the present study was to determine the
12 potential application of spray dried egg yolk powder enriched in bovine RVA-specific IgY
13 Ab as a passive therapeutic treatment to be applied for 7 days after the appearance of
14 diarrhea in neonatal calves experimentally challenged with virulent bovine RVA (G6P[5]
15 strain). The potential of the powder as a commercial product was further evaluated.

16 17 **3. MATERIALS AND METHODS:**

18 **Ethics Statement:** The protocol for animal management and euthanasia met the
19 requirements of The Institutional Animal Care Committee (IACC, protocol number 02-
20 09-08) of the Veterinary College (University of Buenos Aires), the Institutional
21 Committee for Experimental Animals Use and Management (from INTA, protocol
22 number 24/2010 for calves and 20/2011 for chickens).

23
24 **Experimental design:**

1 **Colostrum feeding, experimental groups and passive treatment administration**

2 Fifteen newborn male Holstein calves acquired from the same dairy farm were removed
3 from their dams at birth prior to suckling and moved into isolation facilities within the first
4 4 hours of life. The animals were housed in individual isolation rooms under a strict
5 management protocol as previously described (Parreño et al., 2004). Calves were
6 randomly assigned to one of the following feeding groups (G): **G1**= colostrum (C) + milk
7 supplemented with bovine RVA-specific egg yolk powder with a final ELISA IgY Ab titer
8 to Indiana RVA strain of 4096 (n=4); **G2**= C + RVA Ab free milk (n=5) and **G3**=
9 colostrum deprived (CD) + RVA Ab free milk (n=6). These last two groups of calves
10 were considered controls. Colostrum deprived calves are often found in herds where
11 animals are not artificially assisted and many of the newborns fail to feed before the first
12 hours of life. Calves in G1 and G2 received 1 L of colostrum pool from RVA non-
13 vaccinated cows (but seropositive to RVA due to natural exposure to the virus in the
14 field) within the first 6 hours of life (prior to gut closure). This colostrum had a virus
15 neutralization (VN) Ab titer to Indiana bovine RVA strain (genotype G6P[5]) of 65,536
16 and an IgG1 ELISA Ab titer of 16,384 and was produced as previously described
17 (Parreño et al., 2004, 2010; Vega et al., 2011). All animals in G1 and G2 had a
18 circulating ELISA mean IgG1 Ab titer to Indiana bovine RVA strain of ~1024/4096 in
19 serum after colostrum intake. Calves in G3 did not receive colostrum but 1 L of RVA Ab
20 free milk at this time point, remaining seronegative to bovine RVA, as expected. The
21 milk used in the experiment was commercial sterilized bovine milk for human
22 consumption (Mastellone Hnos., Argentina). All animals were orally inoculated with
23 $10^{5.85}$ Focus Forming Units (FFU) of virulent bovine RVA strain G6P[5] (Indiana strain)
24 between the third and fourth feeding (0 post-inoculation day -PID-) (Fernandez et al.,

1 1998). This viral dose was previously confirmed to cause diarrhea and virus shedding in
2 100% of inoculated CD calves (Parreño et al., 2004). This viral strain is one of the most
3 prevalent RVA strains circulating in newborn calves in Argentina (Garaicoechea et al.,
4 2006; Badaracco et al., 2012) and worldwide (Midgley et al., 2012; Papp et al., 2013).
5 Immediately after the onset of diarrhea, only calves assigned to IgY passive treatment
6 group (G1) received 2 L of the supplemented milk twice a day for 7 days with a final
7 ELISA IgY Ab titer to bovine RVA (Indiana strain) of 4096. After this time, all calves in
8 this group were fed milk without supplemental Abs. Calves in G2 and G3 were fed
9 commercial RVA Ab free milk during the experiment. All the calves were euthanized at
10 PID 21 to study the primary Ab responses to bovine RVA infection.

11
12 **Clinical observations and sample collection:** Calves were examined daily for the
13 development of diarrhea and virus shedding after virulent bovine RVA inoculation.
14 Rectal temperatures were measured during the experiment and values over 39°C were
15 considered as hyperthermia. To estimate the severity of the diarrhea, fecal consistency
16 was scored blindly by qualified technicians as follows: 0: normal; 1: pasty; 2: semi-liquid;
17 3: liquid; considering a score equal or greater than 2 as diarrhea (Parreño et al., 2010).
18 The area under the fecal scores curve (AUC) for each calf was determined and then the
19 average AUC for diarrhea was calculated for each group. Prior and after virulent bovine
20 RVA inoculation, fecal samples were collected daily to assess virus shedding. Serum
21 samples were collected before colostrum feeding (within 4 hours after birth), at bovine
22 RVA inoculation (approximately 40 hours of birth), and then weekly (PID 7, 14, 21).
23 Serum Abs to bovine RVA (Indiana G6P[5] strain) were assayed by ELISA and VN
24 tests. The presence of coproantibodies (coproAb) was also assessed by ELISA.

1 Peripheral blood lymphocytes (PBL) were sampled at PID 0 and then weekly, to assess
2 ASC responses by ELISPOT assay. At PID 21, the animals were euthanized and the
3 number of bovine RVA ASC were quantified by ELISPOT in the following gut-associated
4 lymphoid tissues (GALT): duodenum, jejunum and ileum lamina propria (LP), jejunum
5 and ileum Peyer's patches (PP), jejunum and cecal mesenteric lymph nodes (MLN) and
6 in systemic lymphoid tissues (spleen and PBL). Large (LIC) and small (SIC) intestinal
7 contents from all calves were collected at necropsy for bovine RVA specific Ab detection
8 by ELISA (Parreno, Bejar et al. 2004).

9
10 **Virus:** Virulent bovine RVA (corresponding to Indiana strain, G6P[5] genotype) inoculum
11 used for calf experimental infection consisted of the intestinal contents of a CD calf that
12 was orally inoculated with a fecal suspension containing this RVA strain, as previously
13 described (Fernandez et al., 1998; Parreño et al., 2004). The cell-culture-adapted
14 Indiana RVA strain was also propagated in monkey kidney cells (MA-104) for its use in
15 ELISPOT, ELISA and VN assays (Parreño et al., 2004, 2010).

16
17 **Egg yolk powder:** Lohmann Brown Classic laying hens were immunized
18 intramuscularly in the breasts with live attenuated Indiana RVA strain (genotype G6P[5])
19 containing 10^{10} FFU/ml as previously described (Vega et al., 2011).

20 Collected eggs from immunized hens were grouped weekly and their yolks were diluted
21 in distilled water at a ratio of 1:3 and stored at -20°C. The RVA IgY Ab titer from each
22 pool was determined by Indiana RVA ELISA and VN assays and pools with RVA IgY Ab
23 titers of 16,384 were spray-dried (FlexPump, Galaxie S.A., Argentina) to obtain egg yolk
24 powder. This powder was diluted in the RVA Ab free milk (10 mg/ml) and had an IgY

1 ELISA Ab titer to Indiana RVA strain of 1024. A total of 100 gr of egg yolk powder were
2 diluted in 2 L of RVA Ab free milk to reach an ELISA IgY Ab titer in the supplemented
3 milk of 4096 (VN titer: 2048) to Indiana RVA strain. The product quality and stability
4 were evaluated over a 2 years period. Briefly, samples were kept at room temperature
5 and relative humidity (RH), at 4°C and room RH, at 30°C and 65% RH or at 40°C and
6 75% RH in similar trialuminum packages, to generate data on stability from normal or
7 accelerated conditions, respectively. After 0, 30, 90, 180, 360, 540 and 720 days from
8 powder production, packages were opened and samples were prepared in RVA Ab free
9 milk as noted above. The IgY ELISA Ab titer to Indiana RVA strain was determined
10 immediately. At the same time points, egg yolk powder was sent to a private laboratory
11 to determine the physicochemical composition and microbiological status of the product.

12
13 **Bovine RVA antigen detection:** Virus shedding was detected in fecal samples using
14 an antigen capture ELISA as previously described (Cornaglia et al., 1989). The
15 genotype and molecular identity of RVA was confirmed by nested RT-PCR followed by
16 DNA sequence analysis as described elsewhere (Garaicoechea et al., 2006; Badaracco
17 et al., 2012).

18
19 **Cell culture immunofluorescence assay (CCIF):** Infectious virus titer was assessed
20 by a CCIF assay (Fernandez et al., 1998; Parreño et al., 2010). Fluorescent foci were
21 visualized using FITC-labeled hyperimmune bovine polyclonal antiserum specific to
22 Indiana bovine RVA strain (genotype G6P[5]) prepared at INTA, and fluorescent foci-
23 forming cells were counted using a fluorescence microscope. Titers were expressed as
24 the number of fluorescent focus forming units per ml (FFU/ml). The AUC for each calf

1 was first determined and then the average AUC for virus shedding was calculated for
2 the group.

3
4 **Bovine isotype-specific Ab ELISA:** The IgM, IgA and IgG1 Ab titers to Indiana RVA
5 strain were determined in colostrum, calf sera, feces, LIC and SIC. Specific IgG1 Abs
6 were detected by an indirect ELISA while specific IgM and IgA Abs were measured by
7 capture ELISA using the reagents and protocols previously described (Fernandez et al.,
8 1998; Parreño et al., 2004; Vega et al., 2011).

9
10 **RVA specific IgY Ab ELISA:** The IgY Ab titers to Indiana RVA strain were determined
11 in crude egg yolk pools, calf and hen sera, calf feces and supplemented milks by an
12 indirect ELISA as previously described (Vega et al., 2011).

13
14 **Fluorescent focus reduction virus neutralization (FFN) test:** Virus neutralizing Ab
15 titers to Indiana RVA strain in colostrum pools, egg yolk pools supplemented milk and
16 calf sera were determined by a FFN test as previously described (Parreño et al., 2010;
17 Vega et al., 2011). The VN titer was expressed as the reciprocal of the highest sample
18 dilution that resulted in >80% reduction in the number of fluorescent foci (Tô et al.,
19 1998).

20
21 **Isolation of mononuclear cells (MNC):** Approximately 15 cm of tissue samples of
22 duodenum, jejunum, and ileum lamina propria were collected. Discrete Peyer's
23 patches (PP) (n=4, for each animal) were identified at different points along the
24 mucosal surface of the jejunum and collected individually. A segment of intestine

1 corresponding to a 10 cm portion of the distal ileum continuous PP was also obtained.
2 The mesenteric lymph nodes (MLN) corresponding to the jejunum and ileocecal
3 regions were collected and processed separately. Intestinal tissues were processed as
4 previously described (Vega et al., 2011).

5
6 **ELISPOT assay for bovine RVA-specific ASC:** An ELISPOT assay for quantification
7 of anti-RVA IgM, IgA, IgG1 and IgG2 ASC was conducted to evaluate effector B-cell
8 responses from all calves at PID 21 in several systemic and gut associated (GALT)
9 lymphoid tissues, as previously described (Parreño et al., 2004; Vega et al., 2011).

10
11 **Statistical analysis:** Fisher's exact test was used to compare proportions of calves with
12 diarrhea and virus shedding among groups. The Kruskal-Wallis rank sum (non-
13 parametric) test was used to compare days of onset and duration of diarrhea and virus
14 shedding, cumulative diarrhea scores, AUC, cumulative titers of virus shed and days
15 with elevated rectal temperature, among groups. VN and isotype-specific Ab titers were
16 \log_{10} -transformed prior to statistical analysis.

17 Differences in Ab titers among groups were evaluated by general analysis of variance
18 (ANOVA) followed by DGC (Di Rienzo, Guzmán & Casanoves) multiple comparison test
19 on repeated measures throughout time. Negative samples at a dilution of 1:4 were
20 assigned an arbitrary Ab titer of 2 for the calculation of geometric mean titers (GMTs). At
21 PID 21, the ASC numbers were compared among groups using the Kruskal- Wallis rank
22 sum test. Statistical significance was assessed at $p < 0.05$ for all comparisons. All the
23 statistical analyses were conducted using Infostat® 2011p/19/8/2011 (Argentina)
24 statistical software.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24

3. RESULTS

Passive treatment with egg yolk powder containing bovine RVA IgY Abs significantly reduced diarrhea duration and severity compares with colostrum deprived calves.

For this experiment, newborn calves assigned to each experimental group were orally inoculated with virulent bovine RVA (Indiana strain, G6P[5] genotype) at 36 hours of life. The onset of diarrhea after virus inoculation was statistically similar for all groups of calves, (G1: 2.0 days; G2: 1.2 days; G3: 2 days). In agreement, bovine RVA infectious viral particles were detected in feces from all animals by the same day that diarrhea appeared. Results for diarrhea and virus shedding are detailed in Figure 1. Following the experimental design, calves assigned to G1 received RVA IgY Ab treatment within 2 hours since the development of diarrhea and for seven consecutive days. Diarrhea in this group terminated an average of 3.2 days after the beginning of the passive treatment (range 2 to 5 days). This duration was significantly shorter than the ones registered for CD calves (10.4 days, G3) while animals fed colostrum followed by RVA Ab free milk (G2) developed RVA-associated diarrhea of an intermediate duration (average 7.6 days). Interestingly, virus shedding duration was significantly shorter in calves in G1 (average 4.5 days) than in animals in G2 (average 11.6 days), while calves in G3 showed an intermediate duration of 6 days in average. However, the areas under the curve of virus shedding (AUC) were not significantly different between the

1 experimental groups of calves. Severity of RVA-associated diarrhea, considered as the
2 average AUC of diarrhea score for each group of calves, was also significantly lower in
3 RVA IgY Ab treated calves (G1) than in either of the other groups of animals (G2 and
4 G3). Furthermore, there was a significant reduction in the number of days with RVA-
5 associated hyperthermia for calves in G1 (2.5 days in average) compared with animals
6 in control groups (G2: 12 days and G3: 11.4 days). It is important to mention that
7 animals receiving passive RVA-specific IgY Ab treatment (G1) did not show other
8 clinical signs of bovine RVA infection like anorexia, dehydration or depression (personal
9 observation, data not shown), while they were evident in all calves in G2 and specially in
10 G3.

11 All the calves in G1 cleared the viral infection while they were under passive treatment
12 with IgY Abs and infectious virus was not detected during the experiment (Figure 1a). As
13 shown, IgY Abs to Indiana RVA strain from the passive treatment were detected by
14 ELISA in feces from calves in G1 (Figure 1a). The ELISA IgY Ab titers to RVA varied
15 daily and could be associated with the viral replication and shedding, especially because
16 there was an increase in the titers observed once RVA-infection was cleared, around
17 PID 9-10. Finally, IgY Abs disappeared in one or two days after the end of the passive
18 treatment (Figure 1a).

19

20 **The passive administration of IgY Abs did not suppress the development of**
21 **humoral immune responses to RVA.**

22

23 Passive or active protection against RVA diarrhea is the most desirable outcome of any
24 treatment or vaccination and can be achieved even when continued viral shedding is

1 detected in feces. Moreover, the asymptomatic infection allows development of active
2 immunity to prevent subsequent natural RVA infections. However, we have also
3 previously demonstrated that oral passive treatments based on egg yolk cause a strong
4 positive modulation of the local humoral immune response, and specially the antibody
5 secreting cell responses in the intestinal mucosa (Vega et al., 2011), so further
6 characterization of calves immune responses to RVA were conducted.

7 We studied the local Ab responses to bovine RVA in feces by isotype- specific ELISA.
8 Results obtained are also depicted in Figure 1 together with virus shedding and diarrhea
9 duration. For CD calves in G3, bovine RVA-specific IgM and IgA Abs peaked at PID 6
10 and 7, respectively (Figure 1). A similar pattern was observed in G2 calves (C + RVA Ab
11 free milk), while IgA Ab peaked at PID 8 and IgM Ab showed low titers from PID 5 to 11
12 for G1 animals (C + bovine RVA IgY Ab 4096 milk). It is also evident from Figure 1 that
13 IgG1 Abs to bovine RVA were present around PID 0 in feces from calves receiving
14 colostrum (G1 and G2) while they were absent in G3 calves. However, from PID 8 on,
15 animals in G3 (CD + RVA Ab free milk) showed detectable ELISA titers of Indiana RVA
16 strain specific IgG1 in feces (Figure 1), similarly to the pattern described in serum
17 (Figure 2). By the end of the experiment, all the calves presented IgG1 as well as IgA
18 Ab to Indiana RVA strain in feces. Although no other significant differences were
19 observed between coproAb titers to bovine RVA, calves in G1 showed a trend toward
20 lower RVA Ab titers than animals in G2 and G3 (Figure 1).

21 At PID 21, animals were euthanized and contents from large (LIC) and small (SIC)
22 intestinal sections were collected. Bovine Abs to Indiana RVA strain were also detected
23 by the same ELISA assay as before (see Figure 1). The IgA and IgG1 Abs were
24 predominant, with especially high bovine RVA-specific IgG1 ELISA Ab titer in G3 (CD +

1 RVA Ab free milk) compared to G1 and G2. The IgA Ab titer in SIC from calves in G1
2 was significantly lower than in calves in G3, with G2 showing intermediate responses.
3 All the other Ab titers determined were statistically similar among 3 groups of calves.
4 This was in accordance with the scenario observed in feces at the end of experiment
5 (Figure 1).

6 The profile and isotype of bovine Abs to Indiana RVA strain in serum are depicted in
7 Figure 2. At PID 0 (previous to experimental virulent bovine RVA inoculation), IgG1 Abs
8 to bovine RVA were present only in animals that received colostrum at birth (G1 and
9 G2), indicating its transference from the gut to the bloodstream. This was associated
10 with the presence of VN activity to bovine RVA at this experimental time point for calves
11 in G1 and G2. The IgG1 Ab titers to Indiana RVA strain remained constant in serum
12 samples from all the animals in G1 and G2 during the experiment. In these groups of
13 calves, after virulent bovine RVA inoculation, it was not possible to discriminate if the
14 detected IgG1 in serum was derived from colostrum intake or of it represented active
15 production of IgG1 by the neonates (Figure 2). As expected, while no Abs were detected
16 in serum samples from calves in G3 (CD animals) at PID 0, ELISA Ab titers of IgG1 to
17 bovine RVA increased progressively from PID 7 to PID 21, and they were statistically
18 similar between animals in all experimental groups at PID 14 and until the end of the
19 experiment (PID 21). At PID 21, IgG1 was the main isotype detected in serum, while IgA
20 and IgM were still present in lower titers in all the experimental groups (Figure 2).

21 As expected for a primary immune response against RVA, IgM isotype was detected by
22 ELISA in serum at PID 7 in all calves. At this experimental time point, IgM Ab titers to
23 Indiana RVA strain were significantly higher in G3 than in G1, while G2 showed
24 intermediate responses (Figure 2). The IgM titers peaked around PID 7 and then

1 decreased, reaching statistically similar titers at PID 14 and 21 between groups. On the
2 other hand, IgA Abs to Indiana RVA strain were also detected from PID 7 on in all
3 experimental groups of calves, with a peak around PID 14. Calves in G2 had statistically
4 lower ELISA IgA Ab titers to bovine RVA at PID 14 than animals in G3, while the titers in
5 G1 were intermediate.

6 The VN Abs remained constant in G1 throughout the experiment, while in G3 there was
7 a clear increment from the beginning of the experiment to PID 21. At this experimental
8 time point, VN titers were statistically similar between all groups. Interestingly, animals
9 in G2 showed a decrease in IgG1 Ab titers to Indiana RVA strain from PID 7 to 14
10 (Figure 2).

11
12 **Oral treatment with IgY Abs for 7 days was associated with significantly higher**
13 **ASC responses in GALT at PID 21.**

14
15 At the end of the experiment (PID 21±3), different tissues were collected to determine
16 the number and isotype of Indiana RVA strain-specific ASC by ELISPOT assay. The
17 results obtained for “systemic tissues” (blood, spleen, Jejunal MLN and Ileo-cecal MLN)
18 are depicted in Figure 3 while the ASC responses in GALT are shown in Figure 4. At
19 PID 21, all the animals had ASC to Indiana RVA strain in the studied tissues. In general,
20 the numbers of ASC in systemic tissues of calves in G1 (C + RVA IgY Ab) were lower
21 than in G2 and G3 groups of calves. In particular, the numbers of bovine RVA-specific
22 IgM and IgG1 ASC responses in the spleens of G1 calves (C + RVA IgY Ab) were
23 significantly less than the ones in the G2 and G3 animals. In MLN from jejunal and ileo-
24 cecal regions, most statistical differences were observed in IgG1 ASC to bovine Indiana

1 RVA strain, where colostrum deprived calves in G3 had higher numbers than animals
2 from G1 and G2. Interestingly, in G2 calves (C + RVA Ab free), the predominant ASC
3 isotype detected at PID 21 was IgM followed by IgA in all the systemic tissues studied.
4 In contrast, in G3 calves (CD + RVA Ab free), most of the bovine RVA-specific ASC
5 responses were mainly of the IgG1 isotype in these same tissues (Figure 3).
6 In the case of the GALT responses, most ASC were present in the proximal intestinal
7 lamina propria (duodenum and jejunum) and, in lower magnitude, in PP, independently
8 of the treatment (Figure 4). This finding is in agreement with the site of bovine RVA
9 infection and replication in the small intestine and represents the main GALT effector –
10 duodenum and jejunum- and inductive –ileum- sites, respectively. The ASC responses
11 to Indiana RVA strain in the duodenum were significantly higher in the IgY- treated
12 group (G1) than in the control groups (G2 and G3) for IgM, IgA and IgG2 isotypes
13 (Figure 4). This trend toward higher numbers of bovine RVA ASC was also observed in
14 other tissues, including Jejunum lamina propria and PP. The ASC responses in G1 were
15 mainly associated with IgA and IgG2 isotypes, while in G2 and G3 groups IgG1 and IgA
16 were the most abundant ASC to Indiana RVA strain at PID 21 (Figure 4).

17

18 **Development of an oral treatment based on IgY Abs for RVA treatment in calves.**

19

20 IgY Abs from chicken egg yolk have been demonstrated to be an effective treatment for
21 RVA – associated diarrhea. But further experiments are needed to determine if IgY also
22 could be applied as a commercial product. We studied the stability of egg yolk powder
23 IgY Ab titers to Indiana RVA strain by ELISA in samples kept in trilaminar aluminum
24 packages and kept at different storage conditions. Results obtained from this assay are

1 depicted in Table 1. The avian IgY Abs reacted with Indiana RVA strain until 2 years
2 after powder production if the powder was kept at 4°C or even at room temperature.
3 Under other stability condition, after storing the packaged samples at high temperature
4 (30°C -65% RH and 40°C – 75% RH), IgY Abs showed a progressive loss of activity
5 over time (Table 1).

6 At 180 days (6 months), powder kept at 40°C – 75% RH showed lower ELISA IgY Ab
7 titers to Indiana RVA strain, that continued decreasing until 720 days. Powder kept at
8 30°C – 65% RH showed intermediate stabilities, losing almost all IgY Ab biological
9 activity by the end of the experiment at 720 days (Table 1).

10 The physicochemical composition and microbiological status of egg yolk powder were
11 determined at the packaging stage (0 days from powder production, Table 2) and at the
12 same time points mentioned above. Regarding the physicochemical composition, it
13 remained quite similar until the last check-point (720 days from powder production)
14 when kept at 4°C or even at room temperature. The microbiological status was
15 acceptable until 360 days in samples kept at room temperature, while it extended until
16 720 days for sample storage at 4°C. Further assays should be conducted to determine
17 the safety and commercial viability of a product based on IgY Abs.

18

19 **4. DISCUSSION**

20

21 In the present study we evaluated the use of IgY Abs to bovine RVA as a passive oral
22 therapeutic treatment to control RVA-associated diarrhea in newborn calves. Animals
23 were experimentally inoculated with bovine Indiana RVA strain (genotype G6P[5]), the
24 prevalent RVA strain circulating in the field (Garaicoechea et al., 2006; Badaracco et al.,

1 2012; Midgley et al., 2012; Papp et al., 2013). As control groups, colostrum- deprived
2 animals and calves receiving colostrum, but not treated with any RVA-specific passive
3 treatment were included. Although normally newborn calves would receive at least one
4 liter of colostrum from their mothers, many of them fail to stand in the first hours of life
5 and nurse soon enough before gut closure. These calves are frequently found in dairy
6 herds from Argentina and their survival is compromised by the lack of passively
7 transferred maternal immune factors and antibodies (Tripathi and Vashishtha, 2006;
8 Osaka et al., 2014). The results obtained demonstrate that daily oral administration of
9 egg yolk powder enriched in IgY Ab to bovine RVA at a final ELISA Ab titer of 4096 (VN
10 titer: 2048, to Indiana RVA strain, genotype G6P[5]) significantly reduced the number of
11 days with diarrhea and hyperthermia as well as the severity of the illness when
12 compared with colostrum deprived calves that did not receive any virus-specific
13 treatment.

14 Regarding virus shedding, calves treated with IgY Abs to bovine RVA cleared virus
15 infection before the end of the passive treatment and significantly earlier than control
16 animals receiving colostrum but without IgY Ab treatment (G2). Also, heterologous IgY
17 Abs were detected by ELISA in feces, suggesting that they were not completely
18 degraded in the intestinal tract. The resistance of IgY Abs to the calf's gut environment
19 was also previously observed by our group in the administration of fluid crude egg yolk
20 as a passive prophylactic treatment to prevent RVA-associated diarrhea in newborn
21 calves (Vega et al., 2011).

22 Another important novel feature observed in this study is the induction of a stronger
23 RVA-specific ASC response in calves receiving milk supplemented with chicken egg
24 yolk powder than in control animals receiving colostrum (G2) or CD calves (G3) fed RVA

1 Ab free milk. This feature was previously observed in calves fed milk supplemented with
2 crude egg yolk administered as a preventive strategy for a longer period of time (Vega
3 et al., 2011). These results together demonstrate that this immunomodulatory activity is
4 local, affecting only GALT ASC responses. In the current experiment, calves were fed
5 egg yolk powder for only 7 days, half of the time compared to previous prophylactic
6 experiments. Yet, the modulatory effect of egg yolk supplementation was clearly
7 maintained. These high numbers of bovine RVA-specific ASC detected in the GALT of
8 egg yolk treated animals represent a powerful active immune strategy to control bovine
9 RVA diarrhea as well as potentially other enteric infections. These results are in
10 agreement with the presence of immunomodulatory molecules in the egg yolk, that
11 promote B cell clonal expansion and IgA switching related to anti-inflammatory
12 responses to dietary antigens (Nelson et al., 2007). It has been previously demonstrated
13 that this immunomodulatory effect of egg yolk is not related to IgY Abs itself, as calves
14 treated with semi-purified IgY Abs for 14 days as well as gnotobiotic pigs fed milk
15 supplemented with semi-purified IgY Abs for 9 days did not develop statistically higher
16 ASC responses in GALT than control groups (Vega et al., 2012). However, it seems like
17 IgY Ab treatment caused a partial suppression of active systemic Ab responses to
18 bovine RVA. This could be associated with less severe diarrhea in treated calves, as
19 systemic Ab responses to bovine RVA seem to be higher in colostrum deprived calves,
20 while G2 calves showed intermediate responses.

21 It has been reported that IgY Abs against bovine RVA can be used to prevent virus-
22 associated diarrhea in newborn calves (Vega et al., 2011). In this work we focus on the
23 treatment of clinical signs of bovine RVA infection once they appear, as many farmers
24 would use a specific treatment more than a preventive one. We also shortened the

1 passive treatment from 14 to 7 days when compared to previous prophylactic
2 experiments by our group (Parreño et al., 2010; Vega et al., 2011). However, even with
3 only 7 days of twice daily continuous treatment, IgY Abs were a strong and versatile
4 immune strategy against bovine RVA-associated diarrhea. Another important feature is
5 the fact that the passive treatment administered to calves in this experiment consisted
6 mainly of egg yolk powder, which can be easily manipulated and kept at room
7 temperature for long periods of time (even until 12-16 months). These properties
8 emphasize the advantages of egg yolk as a source of specific antibodies. Avian Ab
9 represent both a reduction and refinement in animal use (Karlsson et al., 2004). When
10 considering Ab production for passive immunotherapy applications, chickens present a
11 much more economical source of large quantities of specific Ab being also less invasive
12 as it only requires daily collection of eggs (Mine and Kovacs-Nolan, 2002; Karlsson et
13 al., 2004; Schade et al., 2005; Kovacs-Nolan and Mine, 2012). It would be critical to
14 develop an egg yolk powder enriched in IgY Abs directed to several enteropathogens
15 involved in neonatal calf diarrhea. This will represent a polyvalent product to be used as
16 a preventive as well as a therapeutic strategy to control calf scours in dairy farms. Being
17 an Ab-based therapy consisting of polyclonal heterologous Abs to bovine RVA, this
18 passive oral treatment will help to control neonatal calf diarrhea. Importantly, this
19 approach will help overcome the problem of microbial resistance to antibiotics, which
20 are used indiscriminately to treat calf diarrhea, offering an innovative, ecological and
21 powerful solution to this serious calf health problem in the dairy industry.

22

23

1 **Acknowledgments**

2 We gratefully acknowledge the technical assistance of José Vallejos, Diego Franco,
3 Nancy Suárez Pérez and Osvaldo Zabal. We would like to thank to Dr. Horacio Terzolo
4 from INTA Balcarce and Pablo Chacana for their advices in IgY technology. We also
5 thank the assistance of “Las Marías” dairy farm, which provided all the calves.

6 Bovine group A Rotavirus strain Indiana (G6P[5]) was kindly provided by Dr. L.J. Saif
7 from Ohio State University (Fernandez et al., 1998).

8 This study was supported by a Basic Biomedical (FIRCABB) (R03), Fogarty Foundation,
9 National Institute of Health (NIH), FIRCA Grant No. 60014561, USA; INTA-BID PICT
10 No. 25375, ANPCyT- Argentina and CVT INTA-Vetanco S.A. N°2934.

11

1 **Figure Legends:**

2 **Figure 1-** Virus shedding, diarrhea duration and coproAbs titers in feces of the
3 experimental groups of calves. Calves in G1 were fed two liters of commercial bovine
4 milk supplemented with 100 g of egg yolk powder containing Indiana RVA strain-specific
5 IgY Abs, twice a day for seven days from diarrhea onset. Calves in G2 and G3 were fed
6 RVA Ab free milk without any supplementation. Continuous lines represent the average
7 ELISA Ab titer of isotype-specific bovine coproAbs to bovine Indiana RVA strain
8 detected in feces. Dashed line in G1 represents the average ELISA IgY Ab titer to
9 Indiana RVA strain. Columns represent average daily profile of virus shedding for
10 experimental calves expressed as FFU/ml. Horizontal bar indicates mean diarrhea
11 duration. Tables included in each graph indicate diarrhea duration defined as the
12 number of days with fecal score ≥ 2 . Feces consistency was scored daily (0=normal;
13 1=pasty; 2=semi-liquid; 3=liquid). ¹ Means in the same column with different superscript
14 upper case letters differ significantly (Kruskal-Wallis rank sum test; $p < 0.05$). ²
15 Determined by ELISA and CCIF. Number of days with bovine RVA-associated
16 hyperthermia was considered as days with body temperature $> 39^{\circ}\text{C}$. C= colostrum / CD
17 = colostrum deprived. AUC: area under the diarrhea/virus shedding curve. PID: post
18 inoculation day. VS: virus shedding. LIC: large intestinal contents. SIC: small intestinal
19 contents.

20

21 **Figure 2-** Geometric mean isotype-specific and neutralizing Ab titers (GMTs) to Indiana
22 RVA (G6P[5] strain) in serum collected weekly (lines). a) G1: calves fed with colostrum
23 (C) followed by bovine RVA egg yolk supplemented milk with a final Indiana RVA IgY Ab
24 titer of 4096 for 7 consecutive days once diarrhea appearance; b) G2: calves receiving

1 C followed by non-supplemented RVA Ab free milk; c) G3: colostrum deprived calves
2 receiving RVA Ab free milk. All calves were orally inoculated with virulent Indiana RVA
3 strain at 0 PID. Statistical analysis of the Ab titers in serum was performed by ANOVA
4 followed by DGC multiple comparison test through time, $p < 0.05$. The “–” indicates no
5 significant differences between groups.

6
7 **Figure 3-** Mean numbers of bovine RVA ASC per 5×10^5 /MNC obtained from systemic
8 lymphoid tissues (PBL, spleen and MLN draining the small and large Intestine), at 21
9 PID. G1: calves fed with colostrum (C) followed by bovine RVA egg yolk supplemented
10 milk with a final bovine RVA IgY Ab titer of 4096 for 7 consecutive days once diarrhea
11 appearance; G2: calves receiving C followed by non-supplemented RVA Ab free milk;
12 G3: colostrum deprived calves receiving RVA Ab free milk. All calves were orally
13 inoculated with virulent Indiana RVA strain at 0 PID. For each tissue, when comparing
14 mean ASC numbers of the same isotype, among treatment groups: different letter
15 indicate a significant difference (Kruskal–Wallis rank sum test, $p < 0.05$). n = number of
16 calves in each group. Error bars indicate SD. The “–” indicates no significant differences
17 between groups.

18
19 **Figure 4-** Mean numbers of bovine RVA ASC per 5×10^5 /MNC obtained from GALT at
20 21 PID. G1: calves fed with colostrum (C) followed by milk supplemented with bovine
21 RVA egg yolk with a final Indiana RVA IgY Ab titer of 4096 for 7 consecutive days after
22 diarrhea appearance; G2: calves receiving C followed by non-supplemented RVA Ab
23 free milk; G3: colostrum deprived calves receiving RVA Ab free milk. All calves were
24 orally inoculated with virulent bovine RVA (Indiana G6P[5] strain) at 0 PID. For each

1 tissue, when comparing mean ASC numbers of the same isotype, among treatment
2 groups: different letter indicate a significant difference (Kruskal–Wallis rank sum test, p
3 < 0.05). n = number of calves in each group. Error bars indicate SD. The “–” indicates
4 no significant differences between groups.

5

6

ACCEPTED MANUSCRIPT

1 **REFERENCES:**

- 2 Badaracco, A., Garaicoechea, L., Rodríguez, D., Uriarte, E.L., Odeón, A., Bilbao, G.,
3 Galarza, R., Abdala, A., Fernandez, F., Parreño, V., 2012. Bovine rotavirus strains
4 circulating in beef and dairy herds in Argentina from 2004 to 2010. *Veterinary*
5 *Microbiology* 158, 394–399.
- 6 Barrandeguy, M., Parreño, V., Lagos Mármol, M., Pont Lezica, F., Rivas, C., Valle, C.,
7 Fernandez, F., 1998. Prevention of rotavirus diarrhoea in foals by parenteral
8 vaccination of the mares: field trial. *Developments in Biological Standardization* 92,
9 253–257.
- 10 Bellinzoni, R.C., Blackhall, J., Terzolo, H.R., Moreira, A.R., Auza, N., Mattion, N.,
11 Micheo, G.L., La Torre, J.L., Scodeller, E.A., 1990. Microbiology of diarrhoea in
12 young beef and dairy calves in Argentina. *Revista Argentina de Microbiología* 22,
13 130–136.
- 14 Bendali, F., Bichet, H., Schelcher, F., Sanaa, M., 1999a. Pattern of diarrhoea in
15 newborn beef calves in south-west France. *Veterinary Research* 30, 61–74.
- 16 Bendali, F., Sanaa, M., Bichet, H., Schelcher, F., 1999b. Risk factors associated with
17 diarrhoea in newborn calves. *Veterinary Research* 30, 509–522.
- 18 Besser, T.E., McGuire, T.C., Gay, C.C., Pritchett, L.C., 1988. Transfer of functional
19 immunoglobulin G (IgG) antibody into the gastrointestinal tract accounts for IgG
20 clearance in calves. *Journal of Virology* 62, 2234–2237.
- 21 Bridger, J.C., 1994. A definition of bovine rotavirus virulence. *Journal of General*
22 *Virology* 75, 2807–2812.
- 23 Cornaglia, E.M., Fitjman, B.M., Schudel, A.A., 1989. Enzyme linked immunoabsorbent
24 assay, immunifluorescent test and electrophoresis analysis of rotaviral RNA in the
25 diagnosis and characterization of the bovine rotavirus. *Revista Latinoamericana de*
26 *Microbiología* 31, 59–62.
- 27 Dhama, K., Chauhan, R.S., Mahendran, M., Malik, S.V.S., 2009. Rotavirus diarrhea in
28 bovines and other domestic animals. *Veterinary Research Communications* 33, 1–
29 23.
- 30 Estes, M., Kapikian, A., 2007. Rotaviruses, in: Knipe, D.M., Howley, P.M., Griffin, D.E.,
31 Lamb, R.A., Martin, M.A., Roizman, B., Straus, S.E. (Eds.), *Fields Virology*.
32 Kluwer/Lippincott, Williams and Wilkins, Philadelphia, PA, pp. 1917–1974.
- 33 Fernandez, F.M., Conner, M.E., Hodgins, D.C., Parwani, A. V., Nielsen, P.R., Crawford,
34 S.E., Estes, M.K., Saif, L.J., 1998. Passive immunity to bovine rotavirus in newborn
35 calves fed colostrum supplements from cows immunized with recombinant SA11

- 1 rotavirus core-like particle (CLP) or virus-like particle (VLP) vaccines. *Vaccine* 16,
2 507–516.
- 3 Garaicoechea, L., Bok, K., Jones, L.R., Combessies, G., Odeón, A., Fernandez, F.,
4 Parreño, V., 2006. Molecular characterization of bovine rotavirus circulating in beef
5 and dairy herds in Argentina during a 10-year period (1994-2003). *Veterinary*
6 *Microbiology* 118, 1–11.
- 7 Hodgins, D.C., Kang, S.Y., DeArriba, L., Parreño, V., Ward, L.A., Yuan, L., To, T., Saif,
8 L.J., 1999. Effects of maternal antibodies on protection and development of
9 antibody responses to human rotavirus in gnotobiotic pigs. *Journal of Virology* 73,
10 186–197.
- 11 Holland, R.E., 1990. Some infectious causes of diarrhea in young farm animals. *Clinical*
12 *Microbiology Reviews* 3, 345–375.
- 13 Ikemori, Y., Kuroki, M., Peralta, R.C., Yokoyama, H., Kodama, Y., 1992. Protection of
14 neonatal calves against fatal enteric colibacillosis by administration of egg yolk
15 powder from hens immunized with K99-piliated enterotoxigenic *Escherichia coli*.
16 *American Journal of Veterinary Research* 53, 2005–2008.
- 17 Ikemori, Y., Ohta, M., Umeda, K., Icatlo, F.C., Kuroki, M., Yokoyama, H., Kodama, Y.,
18 1997. Passive protection of neonatal calves against bovine coronavirus-induced
19 diarrhea by administration of egg yolk or colostrum antibody powder. *Veterinary*
20 *Microbiology* 58, 105–111.
- 21 Karlsson, M., Kollber, H., Larsson, A., 2004. Chicken IgY: utilizing the evolutionary
22 advantage. *World's Poultry Science Journal* 60, 341–348.
- 23 Kohara, J., Tsunemitsu, H., 2000. Correlation between maternal serum antibodies and
24 protection against bovine rotavirus diarrhea in calves. *The Journal of veterinary*
25 *medical science / the Japanese Society of Veterinary Science* 62, 219–221.
- 26 Kovacs-Nolan, J., Mine, Y., 2012. Egg Yolk Antibodies for Passive Immunity. *Annual*
27 *Review of Food Science and Technology* 3, 163–182.
- 28 Kovacs-Nolan, J., Mine, Y., 2004. Avian egg antibodies: basic and potential
29 applications. *Avian and Poultry Biology Reviews* 15, 25–46.
- 30 Kuroki, M., Ohta, M., Ikemori, Y., Icatlo, F.C., Kobayashi, C., Yokoyama, H., Kodama,
31 Y., 1997. Field evaluation of chicken egg yolk immunoglobulins specific for bovine
32 rotavirus in neonatal calves. *Archives of Virology* 142, 843–851.
- 33 Kuroki, M., Ohta, M., Ikemori, Y., Peralta, R.C., Yokoyama, H., Kodama, Y., 1994.
34 Passive protection against bovine rotavirus in calves by specific immunoglobulins
35 from chicken egg yolk. *Archives of Virology* 138, 143–148.

- 1 Lundgren, O., Svensson, L., 2001. Pathogenesis of Rotavirus diarrhea. *Microbes and*
2 *Infection* 3, 1145–1156.
- 3 Malik, S. V., Kumar, A., Bhilegaonkar, K.N., 1995. Rotavirus an emerging
4 enteropathogen of man and animals: an overview. *The Journal of Communicable*
5 *Diseases* 27, 199–207.
- 6 Martella, V., Bányai, K., Matthijssens, J., Buonavoglia, C., Ciarlet, M., 2010. Zoonotic
7 aspects of rotaviruses. *Veterinary Microbiology* 140, 246–255.
- 8 Mebus, C.A., Underdahl, N.R., Rhodes, M.B., Twiehaus, M.J., 1969. Further studies on
9 neonatal calf diarrhea virus. *Proceedings, annual meeting of the United States*
10 *Animal Health Association* 73, 97–9.
- 11 Midgley, S.E., Bányai, K., Buesa, J., Halaihel, N., Hjulsager, C.K., Jakab, F., Kaplon, J.,
12 Larsen, L.E., Monini, M., Poljšak-Prijatelj, M., Pothier, P., Ruggeri, F.M., Steyer, A.,
13 Koopmans, M., Böttiger, B., 2012. Diversity and zoonotic potential of rotaviruses in
14 swine and cattle across Europe. *Veterinary Microbiology* 156, 238–245.
- 15 Mine, Y., Kovacs-Nolan, J., 2002. Chicken egg yolk antibodies as therapeutics in enteric
16 infectious disease: a review. *Journal of Medicinal Food* 5, 159–169.
- 17 Nelson, R., Katayama, S., Mine, Y., Duarte, J., Matar, C., 2007. Immunomodulating
18 effects of egg yolk low lipid peptic digests in a murine model. *Food and Agricultural*
19 *Immunology* 18, 1–15.
- 20 Osaka, I., Matsui, Y., Terada, F., 2014. Effect of the mass of immunoglobulin (Ig)G
21 intake and age at first colostrum feeding on serum IgG concentration in Holstein
22 calves. *Journal of Dairy Science* 97, 6608–12.
- 23 Papp, H., László, B., Jakab, F., Ganesh, B., De Grazia, S., Matthijssens, J., Ciarlet, M.,
24 Martella, V., Bányai, K., 2013. Review of group A rotavirus strains reported in swine
25 and cattle. *Veterinary Microbiology* 165, 190–9.
- 26 Parreño, V., Béjar, C., Vagnozzi, A., Barrandeguy, M., Costantini, V., Craig, M.I., Yuan,
27 L., Hodgins, D., Saif, L., Fernández, F., 2004. Modulation by colostrum-acquired
28 maternal antibodies of systemic and mucosal antibody responses to rotavirus in
29 calves experimentally challenged with bovine rotavirus. *Veterinary Immunology and*
30 *Immunopathology* 100, 7–24.
- 31 Parreño, V., Marcoppido, G., Vega, C., Garaicoechea, L., Rodriguez, D., Saif, L.,
32 Fernández, F., 2010. Milk supplemented with immune colostrum: Protection against
33 rotavirus diarrhea and modulatory effect on the systemic and mucosal antibody
34 responses in calves experimentally challenged with bovine rotavirus. *Veterinary*
35 *Immunology and Immunopathology* 136, 12–27.

- 1 Saif, L.J., Fernandez, F.M., 1996. Group A rotavirus veterinary vaccines. *The Journal of*
2 *Infectious Diseases* 174 Suppl, S98–S106.
- 3 Saif, L.J., Smith, K.L., 1985. Enteric viral infections of calves and passive immunity.
4 *Journal of Dairy Science* 68, 206–228.
- 5 Schade, R., Calzado, E.G., Sarmiento, R., Chacana, P.A., Porankiewicz-Asplund, J.,
6 Terzolo, H.R., 2005. Chicken egg yolk antibodies (IgY-technology): a review of
7 progress in production and use in research and human and veterinary medicine.
8 *Alternatives to laboratory animals : ATLA* 33, 129–154.
- 9 Stott, G.H., Fellah, A., 1983. Colostral immunoglobulin absorption linearly related to
10 concentration for calves. *Journal of Dairy Science* 66, 1319–1328.
- 11 Svennerholm, A.-M., Steele, D., 2004. Microbial-gut interactions in health and disease.
12 Progress in enteric vaccine development. Best practice & research. *Clinical*
13 *gastroenterology* 18, 421–45.
- 14 Tô, T.L., Ward, L.A., Yuan, L., Saif, L.J., 1998. Serum and intestinal isotype antibody
15 responses and correlates of protective immunity to human rotavirus in a gnotobiotic
16 pig model of disease. *Journal of General Virology* 79, 2661–2672.
- 17 Touchette, K.J., Brien, M.L.O., Coalson, J.A., 2003. Liquid Egg as an Alternative Protein
18 Source in Calf Milk Replacers. *Journal of Dairy Science* 86, 2622–2628.
- 19 Tripathi, V., Vashishtha, B., 2006. Bioactive Compounds of Colostrum and Its
20 Application. *Food Reviews International* 22, 225–244.
- 21 Vega, C., Bok, M., Chacana, P., Saif, L., Fernandez, F., Parreño, V., 2011. Egg yolk
22 IgY: Protection against rotavirus induced diarrhea and modulatory effect on the
23 systemic and mucosal antibody responses in newborn calves. *Veterinary*
24 *Immunology and Immunopathology* 142, 156–169.
- 25 Vega, C.G., Bok, M., Vlasova, A.N., Chattha, K.S., Fernández, F.M., Wigdorovitz, A.,
26 Parreño, V.G., Saif, L.J., 2012. IgY antibodies protect against human rotavirus
27 induced diarrhea in the neonatal gnotobiotic piglet disease model. *PLoS ONE* 7,
28 e42788. doi: 10.1371/journal.pone.0042788.

29

30

31

32

1 **Table 1-** Stability of egg yolk powder IgY Ab titer to bovine RVA (Indiana G6P[5] strain)
 2 determined by ELISA in 10 mg/ml solution from samples kept at different storage
 3 conditions.

Storage Conditions	Days from egg yolk powder production						
	0	30	90	180	360	540	720
4°C and room RH	1024	1024	1024	1024	1024	1024	1024
Room Temperature and RH	1024	1024	1024	1024	1024	1024	1024
30°C – 65% RH	1024	1024	1024	256	256	64	4
40°C – 75% RH	1024	1024	1024	256	256	4	4

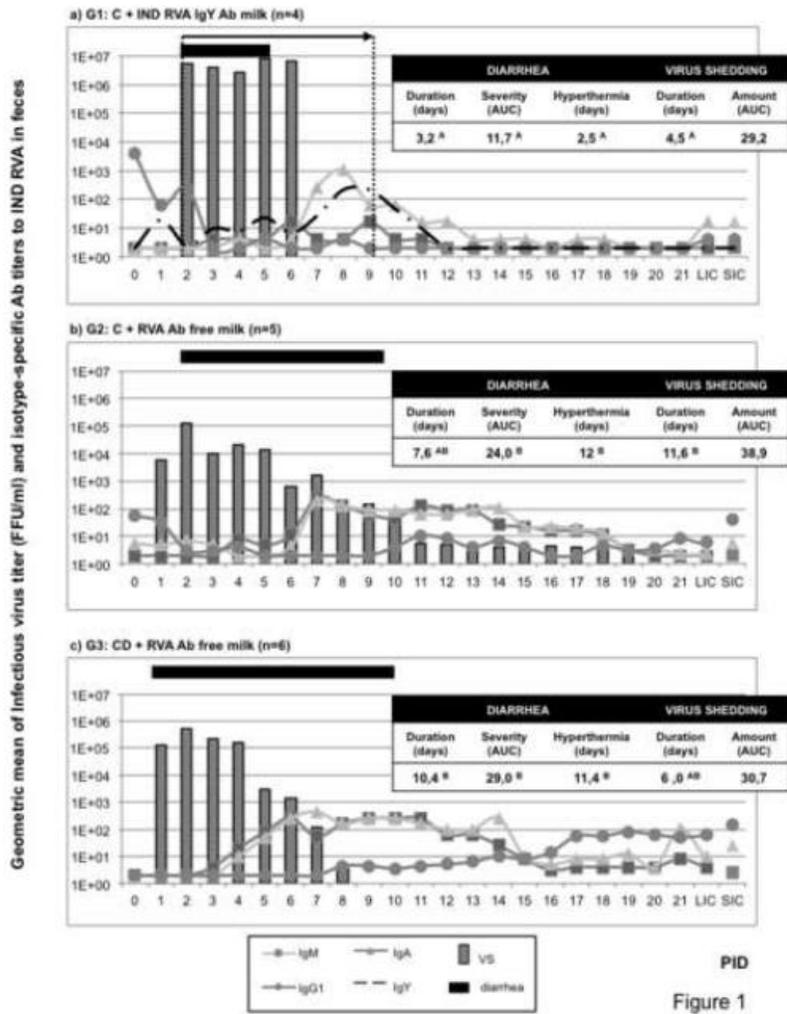
5 After 0, 30, 90, 180, 360, 540 and 720 days from powder production, packages were
 6 opened and samples were prepared in milk as described in Materials and Methods. The
 7 IgY ELISA Ab titer to bovine RVA (Indiana G6P[5] strain) was determined immediately
 8 by ELISA assay. RH: relative humidity.

9 **Table 2-** Physicochemical composition and microbiological status of egg yolk powder at
 10 packaging (0 days from powder production).

Deleted: ¶

Physicochemical composition			Microbiological status	
% Humidity	% Fat	% Protein	Total Coliforms (CFU/g)	<i>Salmonella</i> sp.
2.97	39.63	48.31	<10	Absent (in 25 g)

12 Egg yolk powder samples were specially taken at the packaging stage and sent to a
 13 private laboratory to determine its physicochemical composition and microbiological
 14 status. CFU/g: colony-forming units per gram.



1
2 Fig. 1
3

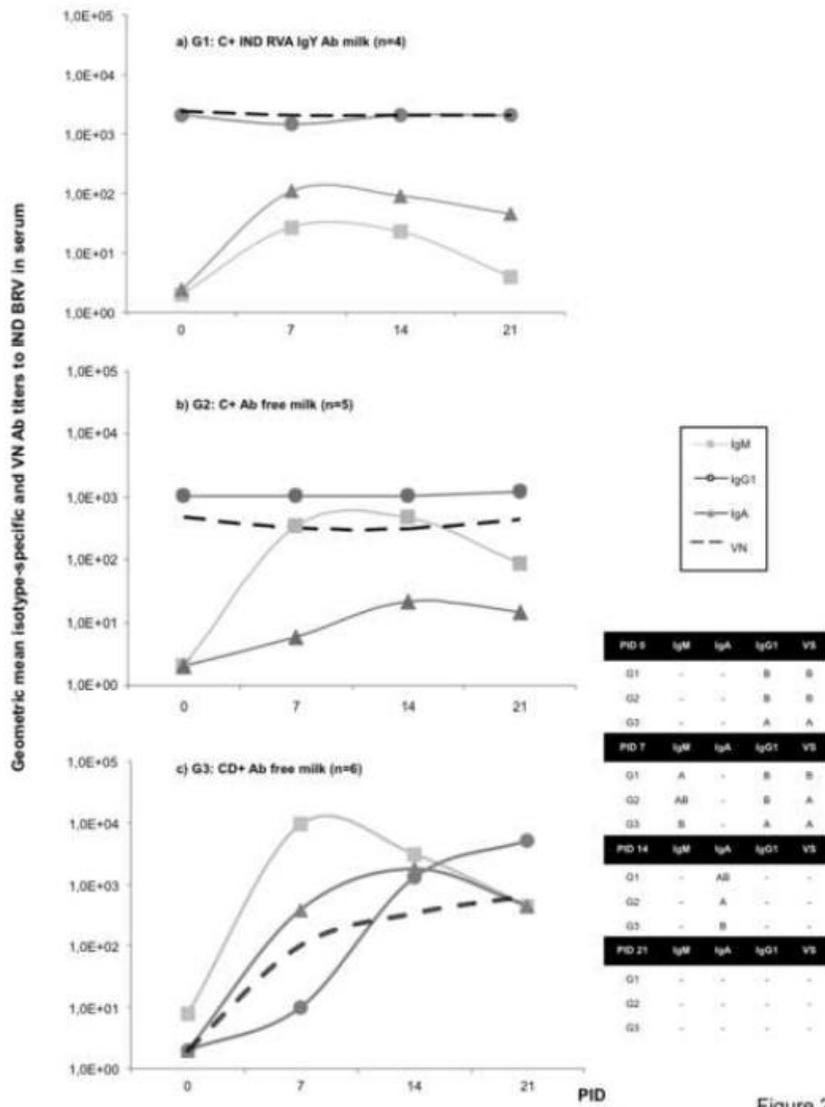


Figure 2

1
2 Fig. 2
3

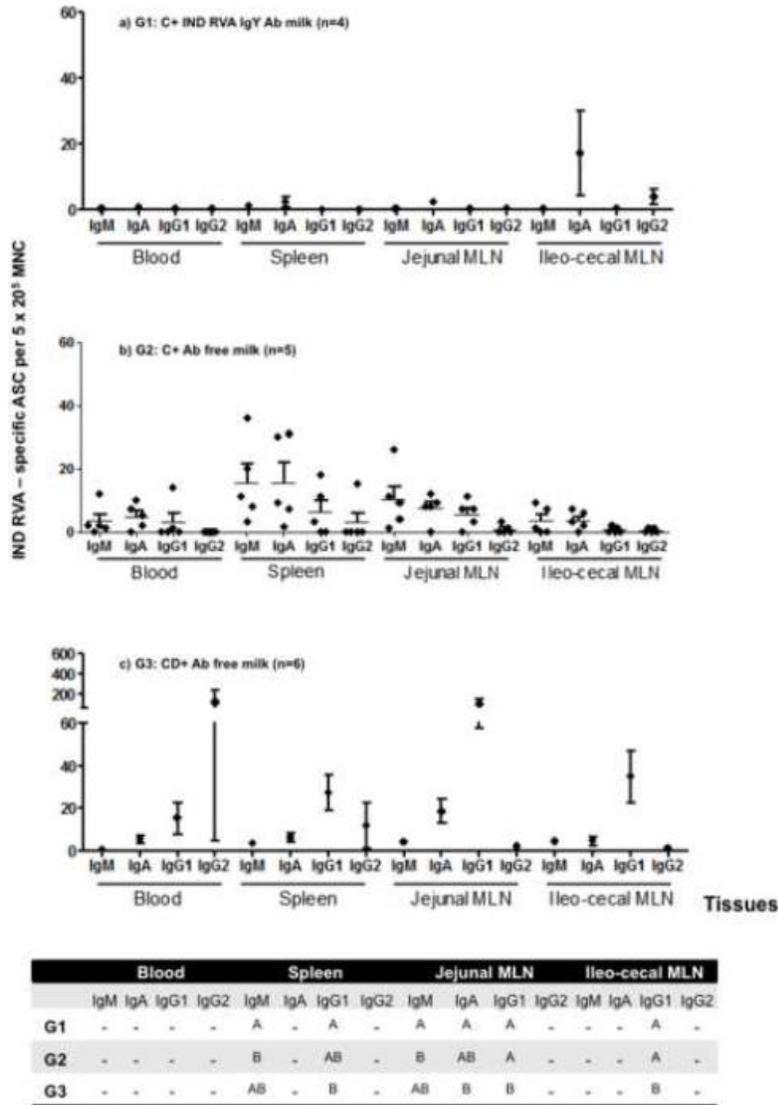


Figure 3

1
2 Fig. 3
3

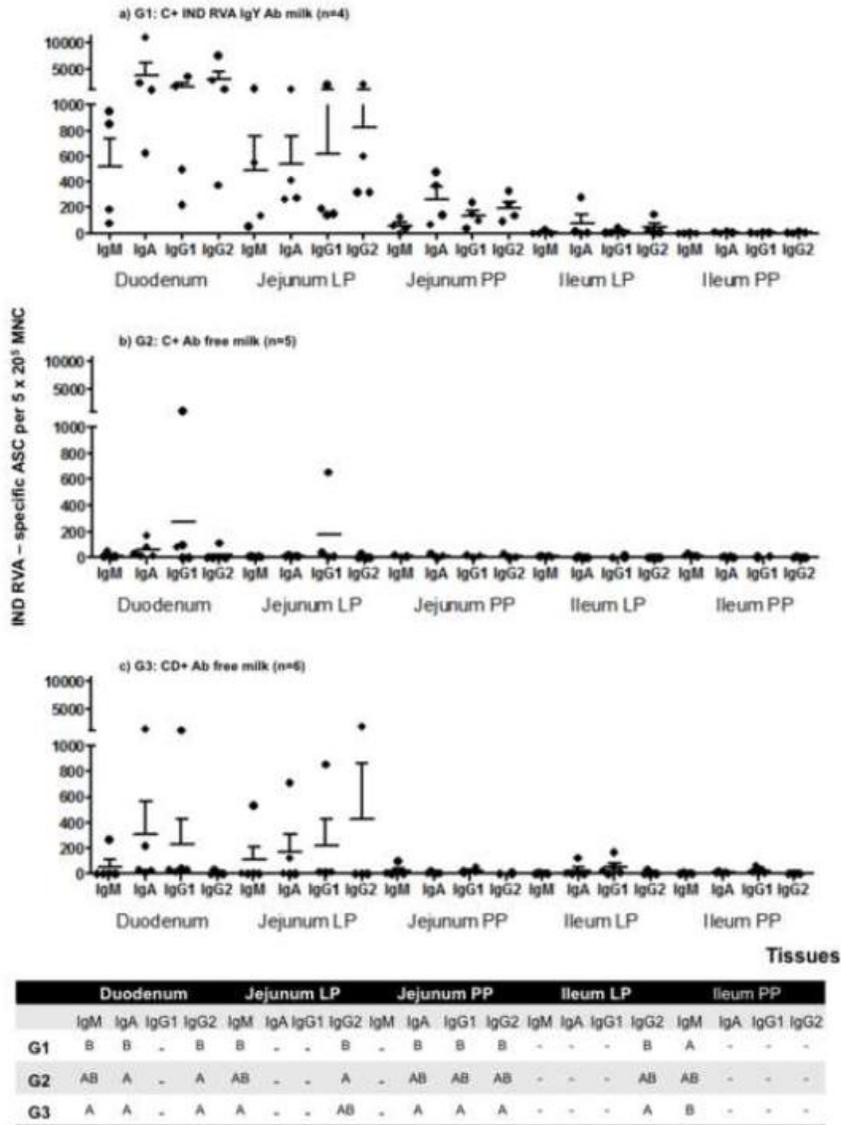


Figure 4

1
2
3

Fig. 4

1
2
3
4
5
6
7
8
9**Highlights**

- Neonatal calf diarrhea is a critical problem and passive therapy with IgY Abs is a way to control it.
- There are no solid studies using Rotavirus specific IgY Abs once calves suffer diarrhea.
- We provide here scientific information regarding the effects of IgY-based products.
- This information is critical considering that IgY Abs are being sold in several countries.
- We prove the therapeutic value of IgY-based treatment and the industrialization of this product.

Formatted: Space After: 10 pt, Line spacing: Multiple 1.15 li, Bulleted + Level: 1 + Aligned at: 0.25" + Indent at: 0.5"

ACCEPTED MANUSCRIPT